

Rapid Agro-Forensic Authentication of Food Products Using FT-IR Spectroscopy and Chemometrics

Dr. Frederick Lia, Karen Attard, Mark Caffari and Malcolm Borg

Abstract

Economic adulteration and geographic mislabelling continue to undermine consumer trust and regulatory integrity in global food markets. As traditional laboratory-based methods remain resource-intensive and time-consuming, Fourier-transform infrared (FT-IR) spectroscopy combined with chemometric analysis has emerged as a rapid, non-destructive, and courtroom-admissible solution. This article reviews recent studies demonstrating the method's effectiveness across diverse food matrices, from honey and olive oil to dairy and coffee. It also explores the growing development of portable FT-IR systems that promise real-time surveillance at border and retail checkpoints.

Introduction

Food fraud, particularly in high-value commodities, is a growing global concern driven by economic motivations (Capuano & Ruth, 2012). It encompasses various practices such as adulteration, mislabelling, and misrepresentation of origin or production methods (Manning & Monaghan, 2019). The complexity of global supply chains has exacerbated this issue, undermining consumer trust and violating food standards (Tola, 2018).

To address such issues, the European Union has implemented quality control schemes such as the Protected Designation of Origin (PDO) and Protected Geographical Indication (PGI). These labels safeguard traditional food products by certifying their geographic origin and production techniques, contributing significantly to rural development and economic value (Sparf, 2010 and Profeta *et al.*, 2009). However, their implementation varies across EU countries, with some nations like Sweden lacking PDO products (Sparf, 2010). Consumer recognition of these labels also differs, influencing purchasing decisions. Italian consumers with excellent knowledge of EU certification labels are more likely to use PDO and PGI logos as their main purchasing motivation (Vecchio & Annunziata, 2011). The increasing global competition has led to cases of food fraud, including the misuse of geographical indications and "Italian sounding" products that mislead consumers (Carreño & Vergano, 2016). This highlights the importance of protecting authentic regional products and educating consumers about these quality labels to maintain their integrity and economic impact.

In parallel, advances in analytical technologies have enhanced the ability to detect and prevent food fraud. Vibrational spectroscopy techniques, particularly FT-IR, NIR, and Raman, have emerged as powerful tools for rapid, non-destructive food analysis (Pandiselvam *et al.*, 2024; Çebi *et al.*, 2023). These methods offer advantages over conventional techniques, including simplicity, adaptability to on-line applications, and absence of hazardous solvents (Pandiselvam *et al.*, 2024). When combined with chemometrics, they enable detailed metabolomic fingerprinting and profiling of food products, allowing for detection and quantification of key constituents (Çebi *et al.*, 2023). FT-IR spectroscopy, in particular, has been extensively used for authentication analysis of fats and oils in food products (Rohman *et al.*, 2020). While mid-infrared spectroscopy is valuable for structural characterization of food components, NIR spectroscopy has gained popularity in the food industry for routine analysis of proximate composition and detection of adulteration (Li-Chan *et al.*, 2006). These techniques offer promising alternatives to more expensive and time-consuming chromatographic and genetic approaches.

FT-IR Spectroscopy and Chemometrics: Principles and Workflow

FT-IR spectroscopy is a powerful analytical technique used to characterize molecular structures and functional groups in various compounds (Siddique, 2024). It operates by measuring the absorption of infrared radiation by molecules, which occurs when the frequency of the radiation matches the vibrational energy of molecular bonds (Álvarez-Ordoñez & Prieto, 2012). The resulting spectrum contains distinct absorption peaks corresponding to specific functional groups, serving as a molecular "fingerprint" (Bates, 1976; Tsuda & Kubouchi, 1993). FTIR spectroscopy can identify various bond types, such as O-H, C-H, and C=O, with each absorbing at characteristic frequencies (Tsuda & Kubouchi, 1993). This technique is widely applicable across scientific domains, including chemistry, biology, and material science, offering insights into molecular interactions, conformational changes, and chemical reactions (Siddique, 2024). Recent advancements, such as attenuated total reflectance (ATR) and micro spectroscopy, have further expanded its utility in both research and industrial applications (Siddique, 2024).

However, raw FT-IR spectra often contain overlapping signals, baseline drift, and scattering effects that obscure meaningful interpretation. To address this, spectral pre-processing techniques are essential. Methods such as Standard Normal Variate (SNV) correction and Savitzky–Golay (SG) derivatives are commonly used to enhance spectral resolution and minimize variability. SNV corrects for scatter-related intensity shifts, while SG derivatives sharpen peaks and highlight subtle spectral features. Optimization of SG parameters,

particularly window size and polynomial order, is critical for effective resolution enhancement (Zimmermann & Kohler, 2013).

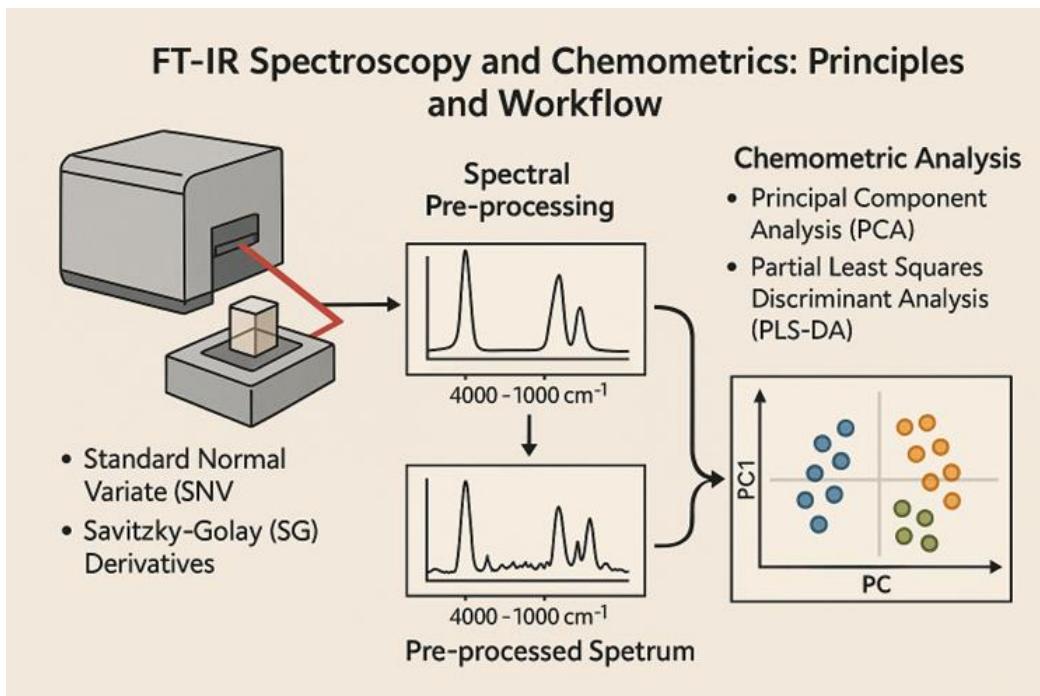


Figure 1: Principles and workflow of FT-IR spectroscopy and chemometrics

When used in combination, these techniques can significantly improve the performance of chemometric models. For instance, SG derivatives paired with Extended Multiplicative Signal Correction (EMSC) have been shown to enhance classification accuracy, although the order of application affects results (Zimmermann & Kohler, 2013). First-derivative SG transformations, for example, have improved the separation of paper types in principal component analysis (PCA) models (Lee *et al.*, 2016). Yet, caution is needed since over-processing can lead to artefacts, especially in datasets with weak signals or limited sample sizes (Delwiche & Reeves, 2010). Moreover, the interaction between SNV and derivative techniques is non-trivial, thus applying them in different sequences can yield different outcomes (Fearn, 2008).

Once spectra are pre-processed, they are typically analysed using multivariate statistical techniques. Unsupervised methods such as PCA are valuable for exploratory analysis and visualizing natural clustering without prior knowledge of sample categories (Sampaio & Calado, 2021; Muehlethaler *et al.*, 2014). In contrast, supervised techniques like Partial Least Squares Discriminant Analysis (PLS-DA) and Soft Independent Modelling of Class Analogy (SIMCA) build predictive models based on known class labels. SIMCA, in particular, has demonstrated high accuracy, sensitivity, and specificity in classifying a variety of materials, including yeast strains (Sampaio & Calado, 2021) and edible salts (Lee *et al.*, 2017), often

outperforming PLS-DA. In spectral data classification, SIMCA achieved correct classification rates of approximately 95%, with notably low false-negative rates (Muehlethaler *et al.*, 2014). To further address issues related to multimodality and nonlinearity in complex spectral datasets, new modeling approaches have been developed. One such method, Nearest Clusters based PLS-DA (NCPLS-DA), has shown improved classification performance compared to conventional PLS-DA, offering better handling of heterogeneous data structures (Song *et al.*, 2018).

Applications Across Food Commodities

Honey:

FT-IR spectroscopy, coupled with chemometric analysis, has proven effective for authenticating honey's botanical and geographical origins. Hennessy *et al.* (2008) applied FT-IR using a germanium ATR crystal and chemometric modelling to classify 150 honey samples by both botanical and geographical origin. Their SIMCA/PLS-DA models achieved 93–100% accuracy, successfully distinguishing Irish and Argentine honeys, among others. Similarly, Ruoff *et al.* (2006) reported error rates as low as <0.1% for certain honey types using ATR-MIR spectroscopy. Additionally, Guyon *et al.* (2021) successfully classified honey varieties based on botanical origin, independent of geographical origin, using SIMCA on ATR-FTIR spectra. These studies collectively support FT-IR as a valuable tool for honey authentication and quality control.

Carob Beans:

Recent studies have explored the use of spectroscopic and chemometric techniques for classifying carob pods and products based on geographical origin and quality parameters. Christou *et al.* (2017) used FT-IR spectroscopy and orthogonal PLS (OPLS) to classify carob pods from seven Mediterranean countries. Spectra focused on the 1200–900 cm⁻¹ region, rich in carbohydrate signals, allowed 100% correct classification by origin, showing promise for PGI applications in carob-derived foods. Kokkinofta *et al.* (2020) further investigated nutritional composition as a marker for provenance, achieving 79% overall classification success using chemometric techniques. Ioannou *et al.* (2023) demonstrated that phenolic profile and antioxidant activity could serve as chemical markers for classifying carob products, with geographical origin significantly affecting these parameters. These studies collectively highlight the potential of spectroscopic and chemometric methods for authenticating carob origin and assessing quality attributes in carob-derived products.

Olive Oil:

Bendini *et al.* (2007) analyzed mono-varietal Italian olive oils using ATR-FT-IR and PCA, achieving clear regional separation based on subtle compositional differences. Similarly, Laouni *et al.* (2022) demonstrated accurate classification of Moroccan olive oils into coastal, continental, and Saharan groups. Similarly, Laouni *et al.* (2022) achieved 84-100% correct classification rates for Moroccan olive oils using FTIR and PCA-LDA/PLS-DA. These studies highlight the potential of spectroscopic methods coupled with chemometrics as rapid, cost-effective tools for verifying olive oil geographical origin and supporting authenticity claims.

Coffee and Dairy:

Wang *et al.* (2011) differentiated Arabica coffees by origin and roast level using FT-IR and SIMCA. The method also detected Robusta adulteration. Moreover, Tarapoulouzi *et al.* (2020) applied FTIR and PCA-OPLS to distinguish cow, goat, and sheep milk cheeses, focusing on bands in the amide I and II regions. Tarapoulouzi & Theocharis (2023) extended this approach to classify traditional Cypriot Halloumi and Anari cheeses by milk species origin. Duliński *et al.* (2025) utilized FTIR spectroscopy with machine learning techniques to authenticate sheep's milk, identifying characteristic spectral signatures, particularly in the amide I and II bands. These studies highlight the potential of FTIR spectroscopy as a rapid and reliable tool for food authentication, with applications in detecting adulterations and ensuring product quality across various food industries

Handheld FT-IR Instruments: Field Applications

Recent studies demonstrate the effectiveness of portable FT-IR devices for on-site food quality control and adulteration detection. Cebi *et al.* (2023) demonstrated that modern portable FT-IR devices equipped with diamond ATR optics and MEMS interferometers offer spectral performance comparable to laboratory instruments. These devices require minimal sample preparation, operate on batteries, and transmit results via wireless connections. Such instruments are capable of detecting adulterants at concentrations as low as 0.4% and achieve R^2 values over 0.98 with root mean square error (RMSE) below 2%.

The versatility of portable spectrometers has been demonstrated across a wide range of food matrices, including milk powder, spices, marine oils, olive oils, honeys, and coffees (Moskowitz & Yakes, 2023). A global study on oregano adulteration found that 33 out of 34 portable near-infrared devices correctly classified 5 out of 6 samples, demonstrating the potential for real-

time remote testing throughout supply chains (McGrath *et al.*, 2020). These portable spectroscopic techniques, combined with chemometrics, offer rapid, accurate, and non-destructive analysis for maintaining food integrity from farm to fork (Çebi *et al.*, 2023).

Limitations and Future Directions

Despite its advantages, FT-IR spectroscopy has some limitations. Variability due to seasonality, climate, or cultivar can affect spectral interpretation. Standardized spectral databases are still under development, and inter-instrument variability can complicate cross-border enforcement.

Moreover, chemometric models require careful calibration and external validation to ensure reproducibility and legal admissibility. Future work should focus on building harmonized spectral libraries and incorporating AI-assisted modelling for improved accuracy and adaptability.

Conclusion

FT-IR spectroscopy combined with chemometrics represents a fast, non-destructive, and scalable approach to agro-forensic analysis. Its successful application across various food commodities highlights its potential as a frontline tool in detecting fraud and verifying origin. With further validation and wider adoption of portable systems, FT-IR could become central to modern food authentication and regulatory enforcement.

Funding

Project 'Authentication of Maltese Pork Meat: Unveiling Insights through ATR-FTIR and Chemometric Analysis' financed by Xjenza Malta through the FUSION: R&I Research Excellence Programme 2024.

References

- Álvarez-Ordoñez, A. and Prieto, M., 2012. Fourier transform infrared spectroscopy in food microbiology. *Journal of Food Science*, 77(2), pp.R14–R22.
- Bates, R.G., 1976. *Determination of pH: theory and practice*. 2nd ed. New York: Wiley.

- Bendini, A., Cerretani, L., Di Virgilio, F., Belloni, P., Bonoli-Carbognin, M. and Lercker, G., 2007. Preliminary evaluation of the application of mid infrared spectroscopy for the classification of virgin olive oils. *Food Chemistry*, 101(2), pp.740–746.
- Capuano, E. and Ruth, S.M., 2012. Fraudulent practices in meat and meat products. *Quality Assurance and Safety of Crops & Foods*, 4(3), pp.141–152.
- Carreño, I. and Vergano, P.A., 2016. Geographical indications and food fraud in the EU: A legal and policy perspective. *Journal of Intellectual Property Law & Practice*, 11(1), pp.15–23.
- Çebi, N., Güler, A. and Öztürk, A., 2023. Advances in FT-IR spectroscopy for food authentication: A review of portable technologies and chemometric models. *Trends in Food Science & Technology*, 135, pp.71–85.
- Christou, E., Kokkinofta, R., Drouza, C. and Agapiou, A., 2017. Geographical discrimination of carobs using FTIR and chemometrics. *Food Chemistry*, 218, pp.284–290.
- Delwiche, S.R. and Reeves, J.B., 2010. Multivariate analysis of near-infrared spectra for classifying forages. *Applied Spectroscopy*, 64(3), pp.327–335.
- Duliński, R., Nowicka, K. and Szpunar-Krok, E., 2025. Authentication of sheep's milk using FTIR and machine learning. *Dairy Science & Technology*.
- Fearn, T., 2008. Pre-processing methods in near infrared spectroscopy. *NIR News*, 19(1), pp.4–6.
- Guyon, C., Chalier, P. and Guillot, X., 2021. Botanical origin classification of honey using FTIR-ATR and chemometrics. *Food Control*, 120, p.107504.
- Hennessy, S., Downey, G. and O'Donnell, C., 2008. Simultaneous analysis of honey for multiple parameters by mid-infrared spectroscopy. *Journal of Agricultural and Food Chemistry*, 56(11), pp.4104–4110.
- Ioannou, I., Christou, E. and Agapiou, A., 2023. Spectral fingerprinting of carob products: phenolic profile and antioxidant markers. *Journal of Food Composition and Analysis*, 114, p.104795.
- Kokkinofta, R., Agapiou, A., Christou, E. and Drouza, C., 2020. Discrimination of carob products by geographical origin using chemometrics. *Food Analytical Methods*, 13(5), pp.1114–1123.
- Laouni, H., El Kamouni, I., El Antari, A. and Mansouri, F., 2022. Authentication of Moroccan olive oils by FT-IR spectroscopy and chemometrics. *Foods*, 11(1), p.38.
- Lee, Y.J., Kim, J.H. and Hwang, H.J., 2016. Comparison of preprocessing techniques in FTIR spectroscopic analysis of food packaging materials. *Analytical Methods*, 8(30), pp.5990–5997.
- Lee, Y.J., Kim, J.H. and Hwang, H.J., 2017. SIMCA-based FTIR classification of edible salts. *Food Chemistry*, 233, pp.41–48.
- Li-Chan, E., Powrie, W.D. and Nakai, S., 2006. The chemistry of food proteins. In: *Food Proteins*. Boston: Springer, pp.3–50.
- Manning, L. and Monaghan, J., 2019. Food fraud: A challenge for the UK. *British Food Journal*, 121(5), pp.1062–1076.
- McGrath, T., Haughey, S.A. and Elliott, C.T., 2020. Real-time detection of oregano adulteration using handheld NIR devices. *Food Chemistry*, 318, p.126483.

- Moskowitz, B. and Yakes, B.J., 2023. Portable spectroscopy for food quality and safety assessment. *Annual Review of Food Science and Technology*, 14, pp.425–446.
- Muehlethaler, C., Leona, M. and Lombardi, J.R., 2014. Classification of FTIR spectra using PCA and SIMCA: a forensic science approach. *Journal of Forensic Sciences*, 59(3), pp.706–713.
- Pandiselvam, R., Manikantan, M.R., Kothakota, A. and Ramesh, S.V., 2024. Applications of FTIR and NIR spectroscopy in food analysis: A review. *Journal of Food Science and Technology*.
- Profeta, A., Balling, R. and Roosen, J., 2009. Consumer preferences and marketing strategies for geographical indication products: Empirical evidence from Europe. *Journal of International Food & Agribusiness Marketing*, 21(3), pp.202–226.
- Rohman, A., Sismindari, S., Erwanto, Y. and Che Man, Y.B., 2020. Authentication analysis of fats and oils by FTIR spectroscopy. *Critical Reviews in Food Science and Nutrition*, 60(5), pp.611–625.
- Ruoff, K., Luginbühl, W., Bogdanov, S., Bosset, J.O. and von der Ohe, K., 2006. Authentication of the botanical origin of honey using mid-infrared spectroscopy. *Journal of Agricultural and Food Chemistry*, 54(18), pp.6867–6872.
- Sampaio, P. and Calado, C.R.C., 2021. Chemometric tools in FTIR-based quality analysis of foods: a comprehensive review. *Food Analytical Methods*, 14(2), pp.235–259.
- Siddique, A., 2024. Fundamentals and advances in FTIR spectroscopy. *Journal of Molecular Spectroscopy*, 391, p.111770.
- Song, W., Kang, H. and Li, Y., 2018. Improved classification of FTIR spectral data using NCPLS-DA. *Chemometrics and Intelligent Laboratory Systems*, 179, pp.99–106.
- Sparf, M., 2010. PDO and PGI in Sweden: Current status and future potential. *European Food and Rural Policy Journal*, 5(2), pp.34–42.
- Tarapoulouzi, M. and Theocharis, S., 2023. Discrimination of Cypriot cheeses using FTIR spectroscopy and chemometrics. *Dairy Science International*, 106(2), pp.157–166.
- Tarapoulouzi, M., Theocharis, S. and Anagnostopoulos, D., 2020. Identification of milk origin in cheeses using FTIR and multivariate analysis. *International Dairy Journal*, 103, p.104612.
- Tola, M., 2018. Global supply chains and food fraud: A critical analysis. *Food Control*, 94, pp.296–304.
- Tsuda, H. and Kubouchi, M., 1993. Analysis of molecular structures using FTIR. *Analytical Chemistry*, 65(7), pp.587A–595A.
- Vecchio, R. and Annunziata, A., 2011. The role of PDO/PGI labelling in Italian consumers' food choices. *Agribusiness*, 27(4), pp.294–313.
- Wang, Y., Tang, J., Johnson, J., Lu, C.D. and Tang, Z., 2011. Coffee authentication using FTIR and SIMCA. *Journal of Food Quality*, 34(3), pp.171–178.
- Zimmermann, B. and Kohler, A., 2013. Preprocessing of vibrational spectroscopy data. *Analytica Chimica Acta*, 794, pp.16–25.